



## Review article

**Bacteriocin: as a promising Bio-preservative for food products**

Garima Verma<sup>\*1</sup>, Rishabh Chitranshi<sup>1</sup>, Anil Pandey<sup>1</sup>, Hem Chandra Pant<sup>2</sup>, Sunil Kumar<sup>3</sup>, Anam Chaudhary<sup>1</sup>, Parul Saini<sup>1</sup>

<sup>1</sup> School of Biological Engineering and Sciences, Shobhit University, Saharanpur, Uttar Pradesh, India

<sup>2</sup> School of Pharmaceutical Science, Jigyasa University (Formerly Himgiri Zee University), Dehradun, Uttarakhand, India

<sup>3</sup> Rajkamal Science and management college, Haridwar, Uttarakhand, India

**Corresponding author:** Garima Verma, ✉ garimav467@gmail.com, **Orcid Id:** <https://orcid.org/0000-0001-8022-1257>

School of Biological Engineering and Sciences, Shobhit University, Saharanpur, Uttar Pradesh, India

© The author(s). This is an open access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by-nc/4.0/>). See <https://jmpas.com/reprints-and-permissions> for full terms and conditions.

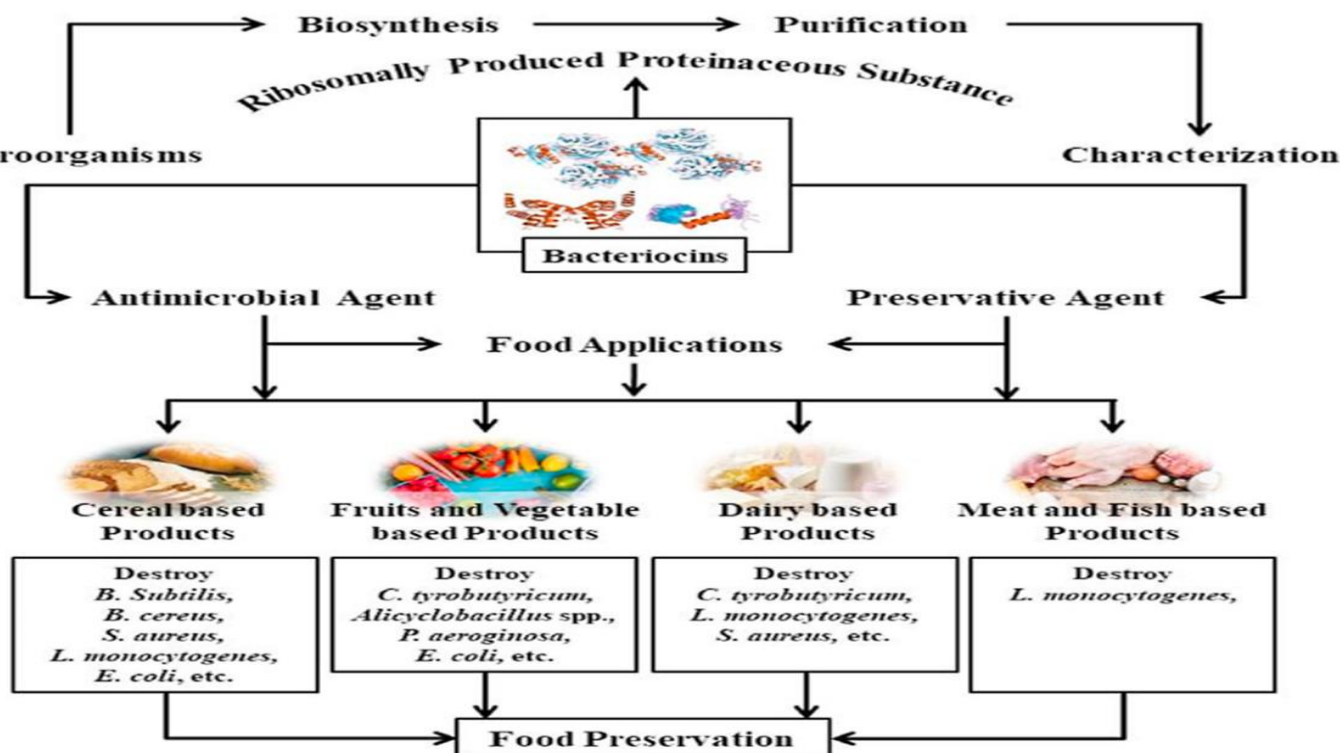
**Received - 11-11-2024, Revised - 27-11-2024, Accepted - 14-12-2024 (DD-MM-YYYY)**

**Refer This Article**

Garima Verma, Rishabh Chitranshi, Anil Pandey, Hem Chandra Pant, Sunil Kumar, Anam Chaudhary, Parul Saini, 2024. Bacteriocin: as a promising bio preservative for food products. Journal of medical pharmaceutical and allied sciences, V 13 - I 6, Pages - 6909 – 6915. Doi: <https://doi.org/10.55522/jmpas.V13I6.6758>.

**ABSTRACT**

Owing to their high nutritional value, high moisture content, and acidic makeup, dairy products are highly perishable. To keep these goods safe and of high quality, they need to be well preserved. Failure to do so results in disease outbreaks and human sickness. These food-borne diseases are major, expensive public health hazards on a global scale. Therefore, proper manufacturing methods, good hygienic standards, etc., are frequently used in the food sector to maintain the quality and safety of goods. There are numerous methods of preservation, starting with low temperature methods such as freezing, refrigeration, etc. and moving up to thermal methods like pasteurization, sterilization, and chemical preservation. The use of contemporary preservation methods including biopreservation, irradiation, and hurdle technology is also widespread today.



A fundamental method of food preservation is bio-preservation, which Utilize of the antibacterial properties of naturally occurring organisms and their metabolites and is able to rationally and in harmony with ancient methods of food preservation and contemporary demands for food safety and quality. The effectiveness of biological antibacterial systems, such as lactic acid bacteria (LAB) and their metabolites such as bacteriocins, is a key component of the biopreservation strategies used to preserve a variety of foods. The amphiphilic properties of some bacteriocin lead to pore development, which disrupts the targeted organisms' cell membranes. As we are all aware, occasionally.

**Keywords:** Bacteriocin, Antimicrobial peptides, Bio-preservation, Food & dairy industries.

## INTRODUCTION

Currently, the shelf-life of a product, which is typically shortened by pathogens, is the main concern in the food sector. Several techniques, including heating, freezing, and chemical preservatives, are utilized to treat these infections. However, recently, bio-preservation techniques, which use bacteriocins specifically, have emerged as a result of advancements in the field of food preservation [25].

Since the times of Robert Koch and Louis Pasteur, environmental microorganisms have been under scientific supervision [39]. In 1929, Alexander Fleming released the antibiotic penicillin, which made it possible to fight particular pathogens [21]. The food industry forbids the use of therapeutic antibiotics, as they pose risks for antibiotic resistance in pathogens [1], they have antibiotic properties and are produced by lactic acid bacteria, bacteriocins such as those produced in the digestive system of humans are not considered true antibiotics. We may easily prevent mistakes between bacteriocins and antibiotics by adhering to these features [9].

Bacteriocins are physiologically active proteins or complex proteins that have antibacterial effects on other bacteria, primarily closely related species [38]. They are bacteriocidal polypeptides produced via ribosomal synthesis [28]. Bacteriocins are a largely varied group that are typically chosen for testing and application as particular antagonists against pathogenic bacteria [46]. The use of bacteriocins can occasionally be restricted for a variety of reasons, and cost is still a significant barrier to employing it more widely [13]. The highly specialized antibacterial protein was referred to as "bacteriocine" by Jacob and others [36].

The term "bacteriocins" refers to a class of antibacterial proteins created by the deadly manufacture of proteins that are adsorbed to particular cell receptors and have a very specific range of function. Later, plasmids were included along with this known relationship with bacteriocin [17]. Since then, the definition has been updated to include the characteristics of bacteriocins made by Gram-positive bacteria. Although there may be a few exceptions, bacteriocins from Gram-positive bacteria typically do not have specialized adsorption receptors, unlike colicins [42]. They have a wider variety of target bacteria and, on average, a lower molecular weight. They also have various models for cell transport and release. Currently, Gram-positive bacterial bactericidal peptides or proteins are

commonly referred to as bacteriocins.

### Background of Bacteriocin

Gratia made the initial discovery of "principev" in 1925. This substance was created when one strain of *E. coli* battled another strain of *E. coli*. Gratia and Fedricq referred to it as a "concine" in 1946, and subsequently in 1953.

### Classification of Bacteriocin

Owing to their origin and unique actions against distinct microorganisms, lactic acid bacteria (LAB) have developed many types of bacteriocins over the years. The majority of bacteriocins are 20–60 amino acid residues long and are cationic, hydrophobic, or amphiphilic compounds [11]. Bacteriocins are primarily grouped into three major classes, with several subgroups, based on their structure, activity, and target mechanisms. A summary of the different kinds of bacteriocins and their sources is provided in Table 1 [32].

#### Class I Bacteriocins (Lantibiotics)

Class I bacteriocins are known as lantibiotics, which are characterized by the presence of unusual amino acids, including lanthionine (Lan), methyllanthionine (MeLan), dehydroalanine, and dehydrobutyrine. These peptides typically have molecular weights less than 5 k Da and contain specific post-translational modifications. There are two types of lantibiotics in Class I: Type A and Type B.

##### Type A lantibiotics

Are elongated peptides that are positively charged? They exert their antimicrobial activity by creating pores in bacterial membranes [15].

##### Type B lantibiotics

Are smaller and often globular peptides that can either be negatively charged or neutral? Their antimicrobial mechanism usually involves the inhibition of enzymatic functions essential for cell wall biosynthesis or other cellular processes.

#### Class II Bacteriocins (Heat-Stable Peptides)

Class II bacteriocins consist of heat-stable, non-lantibiotic peptides, usually with molecular weights less than 10 kDa. These peptides typically consist of standard amino acids and can be further subdivided into three smaller groups [26].

##### Class II A

Bacteriocins, such as pediocin-like peptides, often share a conserved N-terminal sequence (Tyr-Gly-Asn-Gly-Val-Xaa-Cys), which is key to their antimicrobial activity. Pediocin, for instance, is particularly noted for its effectiveness against *Listeria monocytogenes*, making it a popular choice in food preservation [26].

**Class II B**

Bacteriocins include more complex peptides that are less potent in their antimicrobial action compared to Class II A peptides.

**Class II C**

Bacteriocins are small, heat-stable, and secreted via the Sec pathway<sup>[31]</sup>. They are also known for their relatively narrow spectrum of activity.

**Class III Bacteriocins (Large, Heat-Labile Peptides)**

Class III bacteriocins consist of large peptides with molecular weights greater than 30 kDa. These peptides are typically heat-labile and have less utility in food science due to their instability at higher temperatures. Their high molecular weight and structural complexity make them less commonly used in industrial applications compared to Class I and II bacteriocins<sup>[26]</sup>.

Another subclass of bacteriocins, which requires the presence of a protein or carbohydrate moiety to function, has been suggested by some scientists. However, due to their relatively low biochemical activity, these bacteriocins are not typically included in the primary classification groups<sup>[6]</sup>.

**Table 1:** Sources of bacteriocin

| Bacteriocin                        | Source                           |
|------------------------------------|----------------------------------|
| <b>Class I type A Lantibiotics</b> |                                  |
| Nisin                              | <i>Lactococcus lactis</i>        |
| Lactocin                           | <i>Lactobacillus seki</i>        |
| Epidermin                          | <i>Staphylococcus epidermis</i>  |
| Gellidermin                        | <i>Staphylococcus galinarum</i>  |
| Lactacin 481                       | <i>L. lactis</i>                 |
| <b>Class I type B Lantibiotics</b> |                                  |
| Mersacidin                         | <i>Bacillus subtilis</i>         |
| Cinamycin                          | <i>Streptomycescinnamoneus</i>   |
| Ancovecin                          | <i>Streptomyces spp</i>          |
| <b>Class II A</b>                  |                                  |
| Pediocin PA-1/Ach                  | <i>Pediococcus acidi lactici</i> |
| Sakacin A                          | <i>L.sake</i>                    |

**Commercially Available Bacteriocins****Table 2:** Some commercially produced bacteriocins

| Bacteriocin  | Commercial name | Producer Strain        | Manufacture                              |
|--------------|-----------------|------------------------|------------------------------------------|
| Nisin        | Nisaplin        | <i>Lc. Lactis</i>      | Danisco, Copenhagen Denmark              |
| Pediocin PA1 | ALTA 2431       | <i>P. acidilactici</i> | Kerry Biosciences Carrigaline, Ireland   |
| Nisi*n       | MAYNISIN        | <i>Lactococcus sp.</i> | MAYSA GIDA San ve Tic. A.S, Turkey       |
| Nisin        | Nisitrol        | <i>Lactococcus sp.</i> | Bimal Pharma Pvt. Ltd., India            |
| Nisin        | SAFRESH™        | <i>Lactococcus sp.</i> | Wuhan Amth Biotechnology Co. Ltd., China |
| Nisin        | YP-1000 Nisin   | <i>Lc. Lactis</i>      | China (National) Abrasives Corp., China  |

**Application of Bacteriocin in Various Food Industries  
Bacteriocins as Food-bio Preservatives**

The use of bacteriocins as food-biopreservatives may be an effective approach to control food spoilage and food borne pathogenic bacteria. The use of bacteriocin in heat possessed food may reduce the thermal process intensity and both improve the nutritional and organoleptic properties of the food and minimize the heat processing costs. Two relatively recent factor accelerating interest in LAB bacteriocin are the increasing incidence and detection of food-borne diseases and the emerging consumer resistance to highly processed foods<sup>[27]</sup>. Food processors are faced with the dilemma of how to best

Bio-preservatives has to fulfil requirements of being nontoxic, stable, should not affect the organoleptic properties of the food products, economical, accepted by recognized authorities, and used in low concentration for their use on commercial scale<sup>[32]</sup>. Although many other type of bacteriocin such as subtilin, cerein, thuricin, plantaricin etc<sup>[3]</sup>. have been isolated and characterized from different bacteriocin producing strains of bacteria<sup>[8]</sup> yet, till date only commercially produced bacteriocins are nisin (*Lc. lactis*) and pediocin (*P. acidilactici*) and others are still in a process of getting commercial status to be used as 46 food preservatives. Nisin, having broad spectrum of antimicrobial activity, has only been given the GRAS status biopreservatives by Food and Drug Administration and was extremely effective in a number of food systems including canned foods and dairy products to prevent food spoilage and increase shelf life<sup>[10]</sup>. Nisin was found effective antimicrobial against almost all gram-positive bacteria in food. Nisin applications as a food preservative was first evaluated in swiss cheese<sup>[25]</sup> where it effectively prevented blowing (gassing) attributable to the growth of clostridia. Subsequently, nisin use as preservatives was investigated with a large variety of fresh and processed foods. The results obtained by Holla (1990) have indicated that pediocin PA-1 and Pediocin Ach can be used as biopreservation to control both spoilage and pathogenic Gram-positive bacteria in liquid milk, ice cream and cheese. Fowler and Gasson (1991) suggested that a reasonable level of nisin for use in processed cheese is 3.75-12.5 mg/Kg finished products. Different studies have shown that the addition of low concentrations of nisaplin (10-50 mg/Liter) to pasturized milk substantially reduced the bacterial count and increased shelf-life<sup>[17]</sup>.

manufacture attractive minimally prepared food without chemical preservatives and preference for minimally processed foods reflect consumer's perception of what constitutes a natural and healthy food. Food fermented with LAB has a positive image among consumers as healthy and natural substances with yoghurt, dahi and sour-dough based bread being examples.

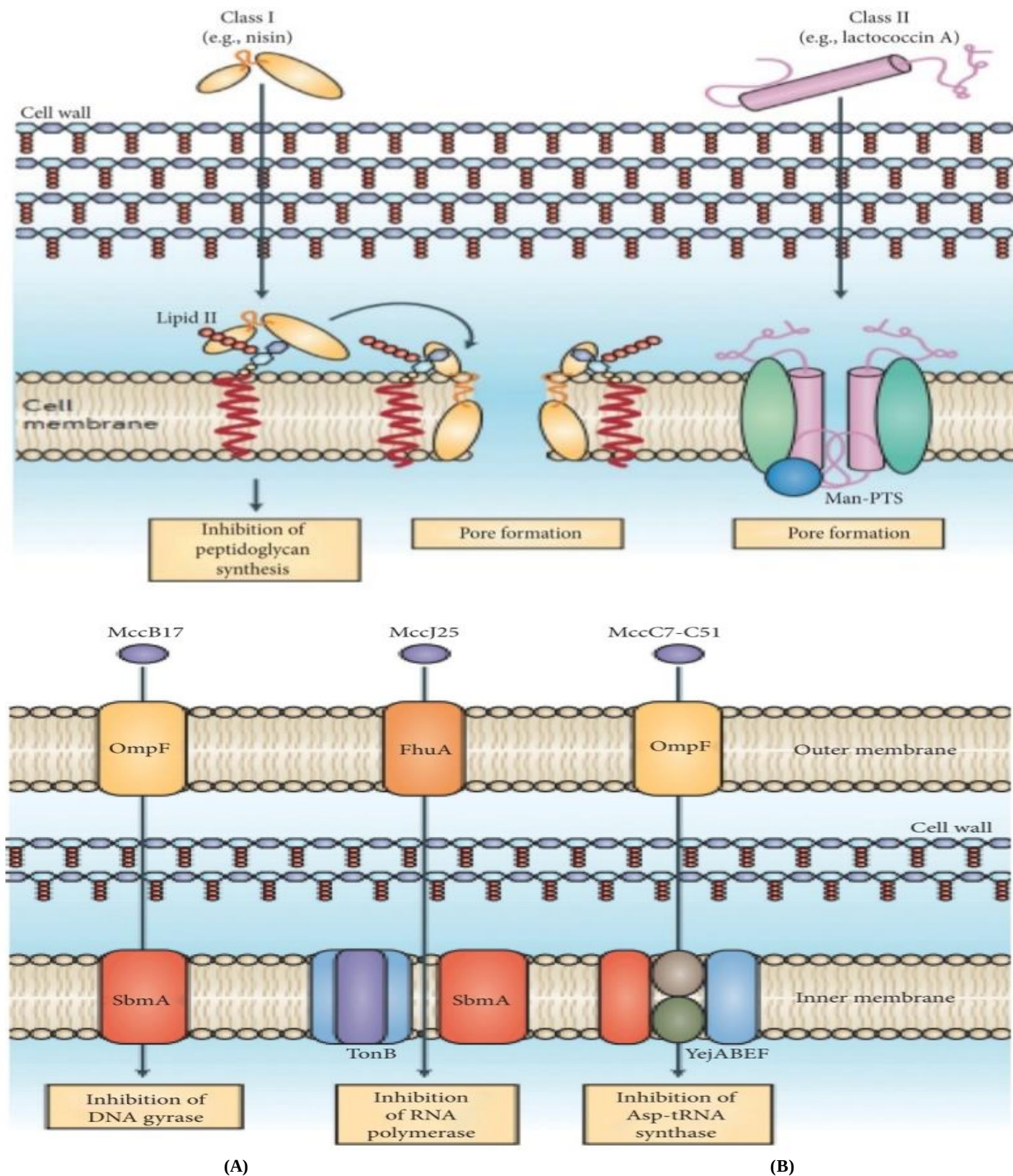
Much recent interest has centred on the potential use of LAB bacteriocin are possible replacement or adjuncts to preservatives that have fallen out of favour with consumers. The safe and efficacious use of nisin for the past 30 years is encouraging and provided the precedent



for continued research on and evaluation of LAB bacteriocin as an emerging class of food preservatives. Recently progress was reported on the on-going EUFAIR project whose aim is to improved quality and safety of agro industrial products by developing novel combination of natural antimicrobial system in conjugation the reduced level of traditional physical and chemical preservation process [20]. It was

reported that test with a novel bacteriocin in model system representing the general composition of dairy products indicated that high levels were required to inhibit *B. cereus* at high abuse temperature an emulsifier was also found to be necessary as fat had a negative effect on bacteriocin activity.

**Figure 1:** (A) & (B) showing mechanism action of bacteriocin of Gram-positive and Gram-negative [16].



### Dairy Industry

Dairy Industry Bacteriocins have wide applications in dairy industry especially during the fermentation of the products. Nisin from *Lc. lactis* and pediocin PA1 produced from *P. acidilactici* PAC1.0 has found their application in dairy industry. Ryan *et al.* (1996) reported the application of lacticin 3147 producing transconjugants to control non-starter Lactic acid bacteria in cheese. Addition of Lactococcin ABM helped in 38 early lysis and release of intracellular enzymes from starter culture and helped in early ripening of Cheddar cheese (O'Sullivan *et al.*, 2002). Use of bacteriocin producing starter cultures was advantageous in protecting fermented foods from transmission of food-borne pathogens as compared to starter cultures lacking bacteriocins. Secondary cultures with bacteriocin producing potential may also contribute to fasten the ripening of dairy products by killing microbes in the primary starter culture. Broad spectrum acidocin CH5 (Chumchalava, 2004), acidocin 1B (KyoungSik *et al.*, 2007) and acidocin AA11 (Abo-Amer, 2007) were reported to have application in fermented products to prevent the growth of pathogenic bacteria. Bacteriocins contributed to the development of flavour and quality of food products. During ripening of cheese, the starter cell culture showed lysis to release proteinases and peptidases which contributed to flavour development in cheese [40].

### Bakery Products and Ingredients

Wheat doughs used for preparation of various bakery products may be contaminated with bacteria resulting in bread defects, including ropiness and enterotoxins production. Bacteriocin at low concentration (14 AU/g) was reported effective against vegetative cells of *B. subtilis*, *B. licheniformis*, *B. cereus* and *B. pumilus* strains under experimental conditions, while its higher concentration (23 AU/g) was required in killing of endospores [42]. *S. aureus* inhibition was also observed with addition of AS-48 in liquid caramel, pumpkin comfiture or diluted almond cream [42]. Bacteriocins were effective against *S. aureus*, *B. cereus* and *L. monocytogenes* in various desserts including baker cream, soy-based desserts and 39 gelatin puddings [42]. *Listeria* contamination has been decreased by >2 log cfu per package by packaging hot dogs with cellulose based films containing 10000 and 7500 AU/mL nisin [24].

### Alcoholic Beverages

Growth of undesirable microorganisms may result in deterioration of the characteristics of alcoholic beverages. Bacteriocins were produced by food-grade lactic acid bacteria and helped in preventing the development of food spoilage bacteria. In the case of wine, it offered the additional advantage of allowing a decrease of sulphurous anhydride levels in wine. Effect of nisin alone and in combination with metabisulphite on red wine indicated measurable reduction of microbial population in the samples with low metabisulphite concentration combined with nisin, thereby, resulting

in the preservation of wine spoilage during the ageing and storage process [40]. A composition prepared by fortifying wort with nutritional supplement, inoculating with nisin producing culture followed by fermentation led to high shelf stability without affecting flavour and aroma of beer.

### Commercially Available Bacteriocins

Biopreservatives has to fulfil requirements of being non-toxic, stable, should not affect the organoleptic properties of the food products, economical, accepted by recognized authorities, and used in low concentration for their use on commercial scale [31]. Although many other type of bacteriocin such as subtilin, cerein, thuricin, plantaricin etc., [4], have been isolated and characterized from different bacteriocin producing strains of bacteria [15] yet, till date only commercially produced bacteriocins are nisin (*Lc. lactis*) and pediocin (*P. acidilactici*) and others are still in a process of getting commercial status to be used as 46 food preservatives. Nisin, having broad spectrum of antimicrobial activity, has only been given the GRAS status biopreservatives by Food and Drug Administration and was extremely effective in a number of food systems including canned foods and dairy products to prevent food spoilage and increase shelf life [5] Nisin was found effective antimicrobial against almost all gram-positive bacteria in food. Nisin applications as a food preservative was first evaluated in swiss cheese [15], where it effectively prevented blowing (gassing) attributable to the growth of clostridia.

Subsequently, nisin use as preservatives was investigated with a large variety of fresh and processed foods. The results obtained by Holla (1990) have indicated that pediocin PA-1 and Pediocin AcH can be used as biopreservation to control both spoilage and pathogenic Gram-positive bacteria in liquid milk, ice cream and cheese. Fowler and Gasson (1991) suggested that a reasonable level of nisin for use in processed cheese is 3.75-12.5 mg/Kg finished products. Different studies have shown that the addition of low concentrations of nisaplin (10-50 mg/Liter) to pasteurized milk substantially reduced the bacterial count and increased shelf-life [22].

**Table 2:** Commercially produced bacteriocins

| Bacteriocin  | Commercial name | Producer Strain        | Manufacture                              |
|--------------|-----------------|------------------------|------------------------------------------|
| Nisin        | Nisaplin        | <i>Lc. Lactis</i>      | Danisco, Copenhagen Denmark              |
| Pediocin PA1 | ALTA 2431       | <i>P. acidilactici</i> | Kerry Biosciences Carrigaline, Ireland   |
| Nisin        | MAYNISIN        | <i>Lactococcus</i> sp. | MAYSA GIDA San ve Tic. A.S, Turkey       |
| Nisin        | Nisitrol        | <i>Lactococcus</i> sp. | Bimal Pharma Pvt. Ltd., India            |
| Nisin        | SAFRESH™        | <i>Lactococcus</i> sp. | Wuhan Amth Biotechnology Co. Ltd., China |
| Nisin        | YP-1000 Nisin   | <i>Lc. Lactis</i>      | China (National) Abrasives Corp., China  |

## CONCLUSION

Currently, the dairy industry's most researched microbial defence strategy is bio-preservation using Bacteriocin. To determine its precise efficacy to particular compounds with antibacterial activity, numerous studies are still ongoing. The quality of biological antimicrobial systems, such as lactic acid bacteria (LAB) and/or their bacteriocins, bacteriophages, and bacteriophage-encoded enzymes, is primarily dependent on the bio-preservation procedures used for different foods. In order to give food products their typical texture and flavor, they are commonly utilized in the food business. They are often used bio-preservatives in the industrialized world and aid in preserving the safety and quality of food. As everyone is aware, chemical preservatives can occasionally have harmful health effects on consumers through a variety of chemical reactions with food and other substances. Occasionally, they can even form carcinogenic chemicals, such as sodium nitrate and sodium nitrite, which are lethal to humans. Thus, contemporary preservation methods such as irradiation, hurdle technologies, and biopreservation are creating a means of releasing humanity from the harmful effects of chemical preservation methods.

## REFERENCES

1. Abo-Amer, A. E, 2007. Characterization of a bacteriocin-like inhibitory substance produced by *Lactobacillus plantarum* isolated from Egyptian home-made yogurt. *Sci. Asia*. 33(3), Pages 313-319. Doi: 10.2306/scienceasia1513-1874.2007.33.313.
2. Aunpad R, Na-Bangchang K, 2007. Pumilicin 4, a novel bacteriocin with anti-MRSA and anti-VRE activity produced by newly isolated bacteria *Bacillus pumilus* strain WAPB4. *Current Microbiology*. 55, Pages 308-313. Doi: 10.1007/s00284-007-0291-y.
3. Bali V, Panesar P S, Bera M B, 2016. Trends in utilization of agro-industrial by-products for production of bacteriocins and their biopreservative applications. *Critical Reviews in Biotechnology*. 36(2), Pages 204-214. DOI: 10.3109/07388551.2014.947250.
4. Chumchalova J, Stiles J, Josephsen J, 2004. Characterization and purification of acidocin CH5, a bacteriocin produced by *Lactobacillus acidophilus* CH5. *Journal of Applied Microbiology*. 96(5), Pages 1082-1089. Doi:10.1111/j.1365-2672.2004.02221.x.
5. Cintas L M, 2001. Bacteriocins: Ecology, genetics, and applications. *FEMS Microbiology Reviews*. 25(1), Pages 44-58. Doi: 10.1016/S0168-6445(00)00057-3.
6. Cintas L M, Herranz C, Hernández P E, et al, 2001. Review: Bacteriocins of lactic acid bacteria. *Food Science and Technology International*. 7(4), Pages 281-305. Doi: 10.1177/108201320100700402.
7. Corr S, C Li, Y Riedel, Cet al, 2007. Bacteriocin production as a mechanism for the antiinfective activity of *Lactobacillus salivarius* UCC118. *Proceedings of the National Academy of Sciences*. 104(18), Pages 7617-7621. DOI: 10.1073/pnas.0700440104.
8. Corsetti A, Hertel C, 2009. Lactic acid bacteria and their bacteriocins: Applications in food preservation. *Food Research International*. 42(7), Pages 1075-1085. DOI: 10.1016/j.foodres.2009.04.010.
9. Corsetti A, Perotti M, 2014. Economic potential of bacteriocins. *Food Research International*. 60, Pages 246-253. DOI: 10.1016/j.foodres.2013.07.029.
10. Cotter P D, Hill C, 2003. Bacteriocins: Developments in their potential as food preservatives. *Journal of Food Protection*. 66(2), Pages 202-206. Doi:10.4315/0362-028X-66.2.202.
11. Paul D Cotter, Colin Hill, R Paul Ross, 2005. Bacteriocins: Developing innate immunity for food. *Nature Reviews Microbiology*. 3(7), Pages 777-788. Doi: 10.1038/nrmicro1213.
12. Paul D Cotter, Colin Hill, R Paul Ross, 2013. Bacteriocins—a viable alternative to antibiotics? *Nature Reviews Microbiology*. 11(2), Pages 95-105. Doi: 10.1038/nrmicro2937.
13. Dufour J P, Laigle A, 1984. Genetic basis of bacteriocin production in Gram-positive bacteria. *Antimicrobial Agents and Chemotherapy*. 8(2), Pages 95-101.
14. Eid R, 2016. Potential antimicrobial activities of probiotic *Lactobacillus* strains isolated from raw milk. *Journal of Probiotics & Health*. 4(2), Pages 138. Doi:10.4172/2329-8901.1000138.
15. Emerenini E C, 2014. In vitro studies on antimicrobial activities of lactic acid bacteria isolated from fresh vegetables for biocontrol of tomato pathogens. *British Microbiology Research Journal*. 4(3), Pages 351. Doi:10.9734/BMRJ/2014/7975.
16. Emerenini F M, Eke C N, 2014. The impact of monetary policy rate on inflation in Nigeria. *Journal of Economics and Sustainable Development*. 5(28), Pages 146-153.
17. Fleming, A. 1929. On the antibacterial action of cultures of *Penicillium*, with special reference to their use in the isolation of *Bacillus influenzae*. *British Journal of Experimental Pathology*. 10(3), Pages 226-236.
18. Flynn S, 2002. Characterization of the genetic locus responsible for the production of ABP-118, a novel bacteriocin produced by *Lactobacillus salivarius* subsp. *salivarius* UCC118. *Microbiology*. 148(4), Pages 973-984. Doi: 10.1099/00221287-148-4-973.
19. Fossi B, 2017, Screening for bacteriocins producing probiotic bacteria from fermented sap of palm trees (*Elaeis guineensis* and *Raffia sudanica*): Production and partial characterization of bacteriocins. *Journal of Applied Biotechnology and Bioengineering*. 2(1), Pages 1-8. Doi: 10.15406/jabb.2017.02.00025.
20. Franklin M N, 2004. Voter turnout and the dynamics of electoral competition in established democracies since 1945. Cambridge University Press. Doi: 10.1017/S1537592705670259.
21. Gänzle M G, 2015. Lactic acid bacteria in food preservation. *Food Research International*. 69, Pages 105-110. Doi: 10.1016/j.foodres.2014.05.035.

22. Gaspar C, 2018. Bacteriocin production of the probiotic *Lactobacillus acidophilus* KS400. *AMB Express*. 8, Pages 1-8. Doi: 10.1186/s13568-018-0674-1.
23. Hoover D G, 2003. Bacteriocins and their food applications." *Comprehensive Reviews in Food Science and Food Safety*. 2(3), Pages 82-100. Doi: 10.1111/j.1541-4337.2003.tb00018.x.
24. Han K S, 2007. Characterization and purification of acidocin 1B, a bacteriocin produced by *Lactobacillus acidophilus* GP1B. *Journal of Microbiology and Biotechnology*. 17(5), Pages 774-783.
25. Holo H, Nes I F, 2000. Lactic acid bacteria: Their bacteriocins and antimicrobial peptides. *Food Research International*. 33(4), Pages 101-109. Doi: 10.1016/S0963-9969(00)00093-9.
26. Holo H. 2002. Bacteriocins of propionic acid bacteria. *Le Lait*. 82(1), Pages 59-68. Doi: 10.1051/lait: 2001009.
27. Ibrahim F, 2020. "Characterization, cytotoxic analysis and action mechanism of antilisterial bacteriocin produced by *Lactobacillus plantarum* isolated from cheddar cheese." *International Journal of Peptide Research and Therapeutics*. 26, Pages 1751-1764. Doi: 10.1007/s10989-019-09946-7.
28. Islam M Z, 2021. Isolation and characterization of dominant lactic acid bacteria from raw goat milk: Assessment of probiotic potential and technological properties. *Small Ruminant Research*. 205, Pages 106532. Doi: 10.1016/j.smallrumres.2021.106532.
29. Jacob H, Rapp J, 1959. Bacteriocins of lactic acid bacteria. *Antimicrobial Agents and Chemotherapy*. 16, Pages 104-107.
30. Kjos M, Langsrud S, 2012. Bacteriocins as food preservatives: Insights into their mode of action. *Journal of Applied Microbiology*. 112(1), Pages 1-11. Doi: 10.1111/j.1365-2672.2011.05119.x.
31. Lizcano L J, Bakkali F, Ruiz-Larrea, et al, 2010. Antioxidant activity and polyphenol content of aqueous extracts from Colombian Amazonian plants with medicinal use. *Food Chemistry*. 119(4), Pages 1566-1570. Doi: 10.1016/j.foodchem.2009.09.038.
32. Lortal S, Chapot-Chartier, M P, 2005. Role, mechanisms and control of lactic acid bacteria lysis in cheese. *International Dairy Journal*. 15(6-9), Pages 857-871. Doi: 10.1016/j.idairyj.2004.08.021.
33. McAuliffe O, Ross R P, 2000. Mechanisms of action of bacteriocins. *Applied Environmental Microbiology*. 66(10), Pages 4236-4242. Doi:10.1128/AEM.66.10.4236-4242.2000.
34. Mokoena M P, 2017. Lactic acid bacteria and their bacteriocins: classification, biosynthesis and applications against uropathogens: a mini-review. *Molecules*. 22(8), Pages 1255. Doi: 10.3390/molecules22081255.
35. Nath S, Chowdhury S, Dora K C, 2015. Application of *Bacillus* sp. as a biopreservative for food preservation. *International Journal of Engineering Research and Applications*. 5, Pages 85-95.
36. O'Sullivan L, Ross R P, Hill C, 2002. Potential of bacteriocin-producing lactic acid bacteria for improvements in food safety and quality. *Biochimie*. 84(5-6), Pages 593-604. Doi: 10.1016/S0300-9084(02)01413-3.
37. O'Connor P M, Mahony J, 2017. Bacteriocins as bio-preservatives. *International Journal of Food Microbiology*. 260, Pages 1-8. Doi: 10.1016/j.ijfoodmicro.2017.08.019.
38. Parvez S, 2017. Role of bacteriocins in food preservation. *Journal of Food Science and Technology*. 54(6), Pages 1117-1125. Doi: 10.1007/s13197-017-2555-9.
39. Parvez S, Kang M J, 2016. Role of bacteriocins in the inhibition of bacterial pathogens. *Biotechnology Advances*. 34(7), Pages 956-969. Doi: 10.1016/j.biotechadv.2016.05.001.
40. Parvez S, 2015. Bacteriocins: A promising and eco-friendly antimicrobial agent in the fight against foodborne pathogens. *Food Control*. 53, Pages 58-71. Doi: 10.1016/j.foodcont.2015.01.023.
41. Riley M A, Wertz J E, 2002. Bacteriocins: Evolution, ecology, and application. *Annual Review of Microbiology*. 56, Pages 117-137. Doi: 10.1146/annurev.micro.56.012302.161024.
42. Sharmila P S, Vidya A K, 2015. Characterization and antibacterial activity of bacteriocin producing *Bacillus subtilis* isolated from raw milk. *International Journal on Applied Bioengineering*. 9(2). Doi: [http://dx.doi.org/ 10.18000/ijabeg.10129](http://dx.doi.org/10.18000/ijabeg.10129).
43. Sorokulova I B, Reva O N, Smirnov V V, et al, 2003. Genetic diversity and involvement in bread spoilage of *Bacillus* strains isolated from flour and rye bread. *Letters in Applied Microbiology*. 37(2), Pages 169-173. Doi: 10.1046/j.1472-765X.2003.01379.x.
44. Surati S, 2020. Bacteriocin, antimicrobial as a new natural food preservative: it's potential and challenges. *Eruditio: Indonesia Journal of Food and Drug Safety*. 1(1), Pages 63-82. Doi: <https://doi.org/10.54384/eruditio.v1i1.34>.
45. Taormina P J, Beuchat L R, 2007. Microbiological Safety of Fresh Produce: A Global Perspective. *Food Control*. 18(6), Pages 303-311. Doi: 10.1016/j.foodcont.2005.07.006.
46. Virk V, Verma G, Ram C, 2022. Production, Purification and Characterization of Bacteriocin Produced by Novel *L. Pentosus* MW857478 for Enhancement of Food Safety and Shelf-Life of Paneer. *Indian Journal of Forensic Medicine & Toxicology*. 16(1). Doi: <https://doi.org/10.37506/ijfmt.v16i1.17396>.