



Research article

Assessment of antimicrobial and *in vitro* antacid activities of Kolakhar: an indigenous herbal soda of Assam, India

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Received – 10 April 2015, Revised – 20 May 2015, Accepted – 25 June 2016 (DD-MM-YYYY)

Refer This Article

Pallab Kalita, Saikat Sena, Chandi Charan Kandarb, 2016. Assessment of antimicrobial and invitro antacid activities of kolakhar: an indigenous herbal soda of Assam, India. Journal of medical pharmaceutical and allied sciences, V 5 - I 4, Pages 356 – 359. Doi: <https://doi.org/10.55522/jmpas.V5I4.0093>.

ABSTRACT

Kolakhar (KK) is a traditionally used soda of Assam. KK is widely used as a detergent or soaps from ancient time to wash cloths and hair. In the rural part of Assam, KK is familiar to treat stomach disorder, respiratory tract disorder and as an antibacterial agents. It is also used as pesticides in different agricultural fields. The present study investigates the antimicrobial and antacid effect of the kolakhar by using disc diffusion method and a modified artificial stomach model. The observation of antimicrobial activity, kolakhar on human pathogenic species using disc diffusion method showed that the kolakhar have more impact on *S. aureus* (21.33 mm). The neutralisation effect, duration of neutralisation effect and capacity were found to be higher for kolakhar than sodium bicarbonate.

Keywords: Cytokines, Polypeptides, Autacoids, Rheumatoid Arthritis, Bone Reabsorption, Tissue Destruction.

INTRODUCTION

Kolakhar is a ethnic herbal soda which is mainly prepared from the banana plant (*Musa balbisiana* colla.). Kolakhar (KK) is a traditional used soda of Assam. KK is widely used as a detergent or soaps from ancient time to wash cloths and hair. Traditionally, it is used as a food additives, especially as a boiling agents. In the rural part of Assam, KK is familiar to treat stomach disorder, respiratory tract disorder and as an antibacterial agents. It is also used as a pesticides in different agricultural fields. Peoples from the different rural areas of Assam, believed that this product also having the antacid activity.

Different parts of banana tree are used to prepare the natural soda commonly known as “kolakhar” in Assam, India. Rhizomes, stem peels are commonly used for this preparation. In general, Kolakhar prepared from Athiya kol (*Musa balbisiana*) was observed to be redish in colour and more dense compared to the Kolakhar obtained from different species of Banana plants. People believed that khar obtaining from athiya kol is more effective than others¹.

A number of studies going on worldwide about the antimicrobial properties of plants. After investigation many have been

used as therapeutic alternatives because of their antimicrobial properties. Plants have many antimicrobial properties as secondary metabolites such as alkaloids, phenolic compounds, etc. In developing countries the practice of complementary and alternative medicine is increase now days on scientific basis for the efficacy of many plants used in folk medicine to treat infections²⁻⁵.

There is a balance between defensive and aggressive factors in normal stomach mucosa. The ulcer etiologies are not yet confirmed but it is generally accepted that peptic ulcers develop when aggressive factors (endogenous, exogenous and/or infectious agents) overcome mucosal defense mechanisms. Some of the main aggressive factors are gastric acid, abnormal motility, pepsin, bile salts, free radicals, use of alcohol and NSAID as well as infection with microorganisms. On the other hand defensive factors such as mucus secretion, bicarbonate production, gastroprotective prostaglandin synthesis, endogenous nitric oxide and normal tissue microcirculation protect against ulcer formation⁶⁻⁷. Recently some approaches to control peptic ulceration include potentiation of the mucosal defense along with reduction of acid secretion and its neutralization, enhancement of antioxidant levels in the stomach,

stimulation of gastric mucin synthesis and inhibition of *H. pylori* growth [7-10]. The currently used drugs produce many adverse effects. Due to the alkalinity nature of kolakhar, traditionally the peoples of Assam are using this product to neutralize the acidity.

Keeping in view the use of kolakhar as an effective home remedy for acidity, the present study was undertaken to evaluate the antimicrobial activity and antacid effect of kolakhar [1].

MATERIALS AND METHODS

Plant Material

The whole plant of *Musa balbisiana* was collected locally from village Ischadagharia, Kamrup, Assam, India. The plant materials were identified and authenticated taxonomically by an expert taxonomist of Guwahati University, Assam, India (Authentication No- 17891RA [1]).

Preparation of Kolakhar

Clean the plant parts with normal water to remove the unwanted materials such as soil. Then slice the different parts in to smaller size and dried in proper sunlight. Then the plant materials was subjected for ignition in open air to convert it to ash. A mixture of 25 gm dry ash of whole plant of *Musa balbisiana* and 500ml of distilled water taken in a one litre conical flask was stirred magnetically for one hour. After filtering, the residue washed with distilled water. The filtrate (light yellow colour) is known as kolakhar. Then the kolakhar is concentrated by evaporation. Dried extracts were kept in refrigerator and used for further study.

Chemicals and Reagents

All the chemicals and reagents were purchased from Sigma Chemical Co., St Louis, MO, USA. Merck Ltd., Mumbai, India. S.D. Fine Chemicals Ltd., Mumbai, India. All other chemicals were obtained from local sources and were of analytical grade.

Instruments

The instruments used in this experiment were a standard pH meter (LABINDIA, SAB 5000), a magnetic stirrer with hot plate temperature controller (IMLH, REMI), an adjustable electrode stand and a peristaltic tubing pump (ELECTROLAB PP 201 V).

Dose Consideration

Doses of kolakhar were taken by considering average animal weight 200gm and dose is 250 mg/kg (kolakhar-I) & 500mg/kg (kolakhar-II).

Assay of Antimicrobial Activity Using Disc Diffusion Method

The 20 ml of sterilized Muller Hinton Agar was poured into sterile petriplates, after solidification, 100 µl of fresh culture of human pathogens were swabbed on the respective plates. The discs were kept over the agar plates using sterile forceps at various concentrations. The plates were incubated for 24 hours at 37°C. After incubation the diameter of inhibitory zones formed around each discs were measured (mm) recorded.

Preparation of Artificial Gastric Acid

Two grams of NaCl and 3.2 mg of pepsin were dissolved in 500 mL distilled water. Hydrochloric acid (7.0 mL) and adequate water

were added to make a 1000 mL solution. The pH of the solution was adjusted to 1.20.

pH Determination of Kolakhar

The pH of kolakhar (kolakhar-I & kolakhar-II) was determined at temperatures ranging from 25°C to 37°C. The pH values of the active control solution sodium bicarbonate (SB) and water was also determined for comparison.

Determination of the Neutralizing Effects on Artificial Gastric Acid

The freshly prepared test solutions kolakhar, water (90 mL) and the active control SB (90 mL) were added separately to the artificial gastric juice (100 mL) at pH 1.2. The pH values were determined to examine the neutralizing effects on artificial gastric juice. (Six experiments were performed for each solution) [2].

Table 1: Antimicrobial Activity of Kolakhar

Pathogen	Drugs	Zone of inhibition (mm)
<i>S. aureus</i>	Distilled water	--
	Standard (ciprofloxacin); 10 µg/disc	25.33±0.23
	Kolakhar; 250 µg/disc	15.67±0.14
	Kolakhar; 500 µg/disc	21.33±0.32
<i>P. aeruginosa</i>	Distilled water	--
	Standard (ciprofloxacin); 10 µg/disc	24.00±0.15
	Kolakhar; 250 µg/disc	16.67±0.26
	Kolakhar; 500 µg/disc	19.33±0.21
<i>C. albicans</i>	Distilled water	--
	Standard (fluconazole); 25 µg/disc	26.33±0.13
	Kolakhar; 250 µg/disc	14.00±0.16
	Kolakhar; 500 µg/disc	18.67±0.18

Determination of The Duration of Consistent Neutralization on Artificial Gastric Acid using the Modified Model of Vatie's Artificial Stomach.

The apparatus of the modified model of Vatie's artificial stomach was made up of three elements: a pH recording system (R), a stomach (S) and a peristaltic pump (P). The stomach was made up of three

portions, S1, S2 and S3. S1 was a reservoir (container), S2 modeled the secretory flux (F-IN), and S3 modeled the gastric emptying flux (F-OUT). Each freshly prepared test sample (90 mL) was added to 100 mL of artificial gastric juice at pH 1.2 in the container of the artificial stomach at 37°C and continuously stirred (30 rpm) with a 2.5-cm magnetic stirring apparatus. Artificial gastric juice at pH 1.2 was pumped at 3 mL/min into the container of the artificial stomach, and pumped out at 3 mL/min at the same time. A pH meter was connected to continuously monitor the changes of pH in the container of the artificial stomach. The duration of the neutralization effect was determined when the pH value returned to its initial value (pH 1.2). Six experiments were performed for each freshly prepared test solution, water and standard (SB).

Determination of the neutralization capacity *in vitro* using the titration method of Fordtran's model

Each freshly prepared test sample (90 mL) was placed in a 250 mL beaker and warmed to 37°C. A magnetic stirrer was continuously run at 30 rpm to imitate the stomach movements. The test samples were titrated with artificial gastric juice to the end point of pH 3. The consumed volume (V) of the artificial gastric juice was measured. The total consumed H⁺ (mmol) was measured as $0.063096 \text{ (mmol/mL)} \times V \text{ (mL)}$. Six experiments were performed for each freshly prepared test solution, water and standard (SB).

Table 2: P^H values with 90 mL water, standard and *kolakhar* added to 100 mL of artificial gastric juice

DRUGS	Hp
Water	1.54 ± 0.02
Standard	2.24 ± 0.16**
Kolakhar-I	1.94 ± 0.06**
Kolakhar-II	1.86 ± 0.02*

Table 3: Duration of antacid effect for consistent neutralization of gastric acid

DRUG	Time(min)
Water	106 ± 4.03
Standard	188 ± 8.67*
Kolakhar-I	174 ± 2.31*
Kolakhar-II	162 ± 7.63*

Data are presented as mean ± SEM (*n* = 6) **P* < 0.01 when compared with water

Table 4. Consumed volume of artificial gastric juice and H⁺ (mmol) in the titration of 90 mL water, standard and *Kolakhar* with artificial gastric juice (pH 1.2) to the end point of pH 3

DRUG	Consumed volume of artificial gastric juice (mL)	mmol of H ⁺
Water	2.42 ± 0.13	0.14 ± 0.006
Standard	70.14 ± 0.118*	3.75 ± 0.08*
Kolakhar-I	20.17 ± 0.75*	1.30 ± 0.05*
Kolakhar-II	19.33 ± 0.92*	1.15 ± 0.06*

Data are presented as mean ± SEM (*n* = 6) **P* < 0.01 when compared with water.

RESULTS

Antimicrobial Activity

Kolakhar showed a good antimicrobial activity against *S. aureus*, *P. aeruginosa* and *C. albicans*. These three different pathogens have tested with commercially available antibiotics and results were

indicated in Table 1. All the test concentrations used against the pathogenic organisms have showed varied degree of antimicrobial activity against the pathogens.

At different temperature (25 to 37), pH values of test samples

Kolakhar prepared from *M. balbisiana* was found to have very high pH (pH 12.0) at temperatures from 25°C to 37°C. At temperatures from 25°C to 37°C, the pH values of water and SB solutions ranged from 6.92 to 7.23 and 9.23 to 9.27, respectively. From this value we can tell that temperature did not affect pH significantly.

Neutralizing capacities of *kolakhar* on artificial gastric acids-

After adding of 90 mL of the test solution with 100 mL of the artificial gastric juice (pH 1.2), the pH values of *kolakhar*-I and *kolakhar*-II solutions were found to be

1.94 ± 0.06 and 1.86 ± 0.02, respectively. The pH values of water and SB solutions were compared with pH values of *kolakhar* at different concentration. This result shows that the neutralizing effect of *kolakhar* was significantly better than that of water.

Duration of consistent neutralization effect on artificial gastric acids-

Neutralizing effects of *kolakhar* solutions were last for 174 ± 2.31 min and 162 ±

7.63 min, respectively. Duration of consistent neutralization effect of water and SB solutions were found 106 and 188 min, respectively. The duration of antacid action of SB was the longest, followed by the *kolakhar* which were significantly higher than that for water [3].

In Vitro Physical neutralization capacity

The consumed volumes of artificial gastric juices to titrate to pH 3.0 for water, *kolakhar*-I, *kolakhar*-II and SB solutions were 2.42, 20.17, 19.33 and 70.14, respectively. The consumed H⁺ were 0.14, 1.30, 1.15 and 3.75 mmol, respectively. The active control SB and both tests exhibited significant antacid potency. Activities. Deregulated production of TNF-α is thought to play a pivotal role in pathogenesis of RA. T-cells are found responsible for the secretion of TNF by direct contact mediated interaction. Histological studies of synovium in RA have indicated that this tissue is very cellular and that several different cell type including macrophages and T-cells are in close proximity. Direct contact mediated interaction using transformed T-cells and monocytes have been found to play a major role in inducing the release of TNF-α, IL10 and metalloproteinase specific surface interaction molecule that are reported to mediate induction of monocyte cytokine synthesis includes CD69, IFA1 CD44 CD40 membrane TNF[29] and signaling lymphocytic activation molecule equilibrium induced by cytokine stimulated T-cell (Tck), monocytes(MQ), monocytes(Tcr) cell receptor dependent stimulated T cells [4].

DISCUSSION

Infectious diseases are the major cause of morbidity and mortality worldwide. The number of multidrug resistant microbial strains and the appearance of strains which reduced susceptibility to antibiotics are continuously increasing. Such increase has been attributed to indiscriminate use of broad spectrum antibiotics, immunosuppressive agents, intravenous catheters organ transplantation and ongoing epidermis of human immunodeficiency virus (HIV) infections. This situation provided the impetus to the search for new antimicrobial substances from various source like medicinal plants. Stomach is an organ which undergoes propulsion, mixing of food, digestion and absorption of food along with the secretory functions. The parietal cells of the stomach secrete about 2500mL of the gastric juice daily. The acid in this gastric juice kills many bacteria and provide a low pH for pepsin to start protein digestion. Mucosal erosions or ulcerations take place when aggressive factors overwhelm the defensive factors of the gastrointestinal mucosa. This leads to the arrival of gastritis, peptic ulcer and gastro oesophageal reflux disease. The main aggressive factors well established for several decades are acid and pepsin. Hence peptic ulcer diseases are mostly treated with antacids, H₂ receptor antagonists and proton pump inhibitors [5].

In this present study, preliminary screening for antimicrobial activity showed that the *kolakhar* exhibited maximum inhibitory zone (30 mm) against *Staphylococcus* and *kolakhar* showing the good potential as an antacid. In our laboratory *in vitro* study also found the considerable antacid activity of *kolakhar*. With more investigation and proper scientific formulation of *kolakhar* may be give a good herbal antacid and anti microbial agents [6].

ACKNOWLEDGEMENT

Authors are thankful to Assam down town University, Guwahati for providing necessary infrastructure to carry out this work. We also acknowledge Dr. Biplab Kumar Dey, Principal, Dept. of

Pharmacy, Adu for his support. This work is a part of Ph.D research work of Mr. Pallab Kalita under the guidance of Dr. Saikat Sen and Dr. C.C. Kandar registered under Assam down town University, Guwahati, Assam.

REFERENCES

1. Deka DC, Talukdar NN, 2007. Chemical and spectroscopic investigation of kolakhar and its commercial importance. Indian J Trad Knowledge. 6(1), Pages 72-78.
2. IE Oboh, JO Akerele, O Obasuyi, 2007. Antimicrobial activity of the ethanol extract of the aerial parts of sida acuta burm.f. (malvaceae). Tropical Journal of Pharmaceutical Research. 6 (4), Pages 809-813.
3. Rajesh Kowti, Harsha. R, Mohammed Gulzar Ahmed, Hareesh AR, 2010. Antimicrobial activity of ethanol extract of leaf and flower of Spathodea campanulata P. Beauv. RJPBCS. 1(3), Pages 691-698.
4. Wu J.Z Cheng, C C Shen, L.L Wang, antioxidant ability on H₂O₂-induced apoptosis in PC12 cells. Int. J. Mol. Sci., Pages 18525–18539.
5. Ford ES, Giles WH, Dietz WH, 2002. Prevalence of metabolic syndrome among US adults: findings from the third National Health and Nutrition Examination Survey. JAMA. 287(3), Pages 356–359. Doi:10.1001/jama.287.3.356.
6. Sandhya S Gouthami gade, Vinod kombath ravindranvidya SE, 2012. Invitro and invivo evaluation of the inhibitory effect of limnophila indica (linn.) druce on shigellosis.journal of Chinese integrative medicine. 10(5), Pages 538-545.