



## Research article

## GC-MS analysis and antimicrobial activity of fixed oil from Saudi *Nigella sativa* (Ranunculaceae)

Abdel Karim M\*, Sufian A, Inas O

<sup>1</sup> Department of Chemistry, Faculty of Science, Sudan University of Science and Technology, Sudan

<sup>2</sup> Department of Chemistry, Faculty of Science, Taibah University, Sudan

<sup>3</sup> Faculty of Applied and Industrial Sciences, University of Bahri, Sudan

Corresponding author: Abdel Karim M, ✉

Department of Chemistry, Faculty of Science, Sudan University of Science and Technology, Sudan

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### ABSTRACT

Constituents of *Nigella sativa* seed oil were identified by GC-MS analysis. Major constituents are: methyl 12-hydroxy-9-octadecenoate (17.37%), 9,12,15-octadecatrienoic acid (14.98%), 9-octadecenoic acid methyl ester (12.84%),  $\beta$ -sitosterol (11.95%), 9,12-octadecadienoic acid methyl ester (10.15%), 11-eicosenoic acid (9.67%), hexadecanoic acid (8.63%), 13-docosenoic acid (5.20%). The oil was extracted by two different methods (soxhlet and maceration) and each sample was screened separately for antimicrobial activity against a panel of pathogens.

**Keywords:** *Nigella sativa*, Fixed oil, GC-MS, Antimicrobial activity.

### INTRODUCTION

*Nigella sativa* (Ranunculaceae) is a small herb about 45cm in height. The plant is cultivated in many countries for its economic value. This plant is widely used in ethnomedicine to treat a wide array of human disorders. The popularity of this herb in Islamic communities is probably due to the ideological belief that the herb is a remedy for multiple diseases. It is used as astringent, stimulant, diuretic, emmenagogue and anthelmintic. It is also used for fever, dyspepsia, piles and skin diseases. Seed oil is used as a local anaesthetic. The alcoholic fraction constitutes about 20%(w/w); aqueous extract 15%(w/w); fixed oil 25-32% (w/w); volatile oil 0.42%(w/w). Seeds contain, among others, nigellone; negellidine; negellimine; steroids; terpenoids; tannins; oleic acid; linoleic acid; saponins; protein; reducing sugar and bitter principle.

Studies on pancreatic cancer cells testify that thymoquinone- a major constituent of seed oil- is synergic with gemcitabine and oxaliplatin. Thymoquinone was also found to be cytotoxic for several types of human cancer cells. Also it was reported that thymoquinone inhibited benzopyrene – induced carcinogenesis in model animals. Some thymoquinone conjugates were found to be active against some resistant tumor cells. A combined dose of selenium and thymoquinone

resulted in decreased cell count, decreased alkaline phosphatase level and decreased glutathione level on the proliferation of osteoblast cells (MG 63). Thymoquinone was found to inhibit tumor incidence and tumour burden significantly in 20-methylcholanthrene-induced fibrosarcoma. It was demonstrated that it protects rats against

NAME-induced hypertension and renal damage are probably by its antioxidant potential.

The essential oil of *Nigella sativa* and the ethyl acetate fraction were cytotoxic against P815 cell line. A decoction comprising *Nigella sativa* seeds, *Hemidesmus indicus* root bark and *Smilax glabra* rhizomes is said to inhibit DEN-mediated expression of GST-P. It also inhibited histopathological changes leading to tumor development in model animals. In vitro studies testified that the ethanol extract of seeds inhibited cancer cells and endothelial cell progression, while in vivo studies demonstrated that topical application of seed extract inhibited skin carcinogenesis. The aqueous and ethanol extracts inactivated MCF-7 breast cancer.

Aqueous extract of seeds inhibited electrogenic intestinal absorption of glucose in vitro. After chronic oral administration, seed extract improved glucose tolerance and body weight in model

animals[23] . Such findings validate the traditional use of *Negella sativa* seeds against diabetes. The ethanol extract of seeds significantly reduced elevated blood glucose, plasma insulin and improved antioxidant enzymes like superoxide dismutase. Furthermore, it reduces glutathione and glutathione peroxidase in the liver and kidney [1].

## MATERIALS AND METHODOLOGY

### Materials Plant material

*Negella sativa* seeds were purchased from the local market, Reyad, Saudi Arabia, and authenticated by Botanist- Dr Mohamed Abhery-Biotechnology

Laboratory, Faculty of Science, Taiba University, Saudi Arabia.

### Extraction and esterification of oil

Powdered *Nigella sativa* seeds (400g) were extracted with n-hexane (Soxhlet). The solvent was removed in vacuo to afford the oil . The extracted oil was esterified by a methanolic solution of sodium hydroxide and methanolic sulphuric acid. After the usual workup, the ester solution (1 $\mu$ l) was directly injected in the GC-MS vial

### GC-MS analysis

For GC-MS analysis, a Shimadzu GC-MS-QP2010 Ultra instrument with an RTX-5MS column (30m, length; 0.25mm diameter; 0.25  $\mu$ m, thickness) was used. Oven temperature program:

Rate : -- ; Temperature oC: 150.0; Hold time (Min-1): 1.00

Rate: 4.00; Temperature oC : 300.0; Hold time (Min-1): 0.00

A Tabulation of chromatographic conditions is shown below [2].

|                         |             |
|-------------------------|-------------|
| Column oven temperature | 150.0°C     |
| Injection temperature   | 300.0°C     |
| Injection mode          | Split       |
| Flow control mode       | Linear      |
| Pressure                | 139.3KPa    |
| Total flow              | 50.0ml/ min |
| Column flow             | 1.54ml/sec. |
| Linear velocity         | 47.2cm/sec. |
| Purge flow              | 3.0ml/min.  |
| Spilt ratio             | - 1.0       |

### Antimicrobial assay

#### Formulation

Aliquots(1ml) of a 24-hour broth culture of the test organisms were aseptically distributed onto nutrient agar slopes and incubated at 37°C for 24 hours. Bacterial growth was harvested and washed off with sterile normal saline, and then suspended in normal saline(100ml) giving about 10<sup>8</sup>-10<sup>9</sup>colony forming units per ml. The

suspension was kept at 4°C until used. Average number of viable organism per ml was assessed by surface viable counting technique.

Serial dilutions of the stock suspension were made in sterile normal saline. Volumes (0.02 ml) of the appropriate dilutions were transferred into the surface of dry nutrient agar plates. Plates were left at room temperature for the drop to dry, and then incubated at 37°C for 24 hours.

Fungal cultures were prepared on potato dextrose agar and incubation was continued for four days at 25 °C. Fungal growth was harvested, washed with sterile normal saline and stored in the refrigerator until used.

### Testing for antibacterial activity

Sterile molten nutrient agar(200ml) was heated at 45°C in a water bath. Bacterial stock suspension (2ml) was added to the agar and mixed. Aliquots(20ml) of the incubated nutrient agar were distributed into sterile Petri dishes. The Petri dishes were divided into two halves (each half was assigned for a test solution). Two cups in each half (10 mm in diameter) were cut by a cork borer (No 4). Positive controls - standard antimicrobial chemotherapeutics-were assigned separate Petri dishes.

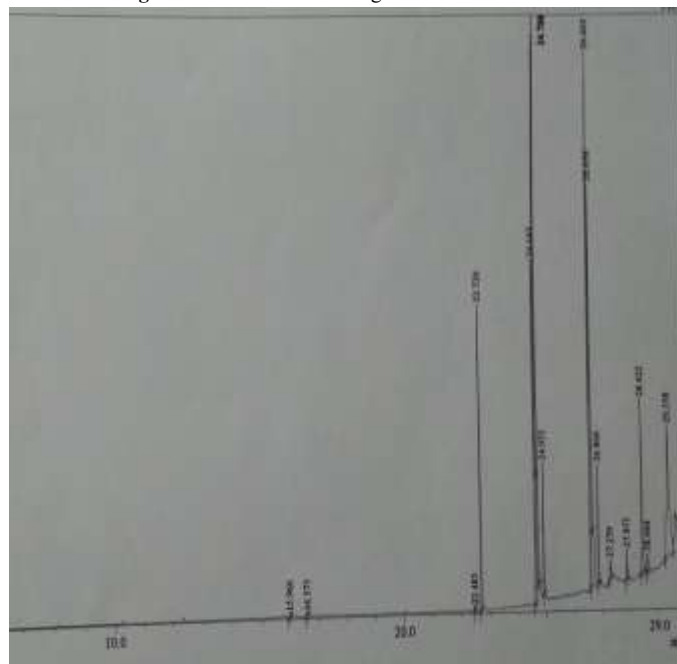
Alternate cups were filled with 0.1 ml samples and allowed to diffuse at room temperature for two hours and then incubated at 37°C for 24 hours. Following incubation, the diameters of the growth inhibition zones were measured in triplicate and averaged.

## RESULTS

### Identification of oil constituents

Sixteen components were detected by GC-MS(Table1). A typical total ion chromatogram (TIC) is shown in Fig.1.

**Figure 1:** Total ion chromatograms of hexane extract



**Table 1:** Constituents of *Negella sativa* seed oil

| Peak# | R.Time | Area      | Area%  | Name                                       |
|-------|--------|-----------|--------|--|
| 1     | 15.966 | 181695    | 0.16   | 1s,4R,7R,11R-1,3,4,7-Tetramethyltricyclo   |
| 2     | 16.575 | 177698    | 0.16   | Butylated Hydroxytoluene                   |
| 3     | 22.485 | 161356    | 0.14   | 9-Hexadecenoic acid, methyl ester, (Z)-    |
| 4     | 22.720 | 9747365   | 8.63   | Hexadecanoic acid, methyl ester            |
| 5     | 24.643 | 11461904  | 10.15  | 9,12-Octadecadienoic acid (Z,Z)-, methyl e |
| 6     | 24.702 | 14494959  | 12.84  | 9-Octadecenoic acid (Z)-, methyl ester     |
| 7     | 24.726 | 16915744  | 14.98  | 9,12,15-Octadecatrienoic acid, methyl este |
| 8     | 24.933 | 3749464   | 3.32   | Methyl stearate                            |
| 9     | 26.602 | 19619456  | 17.37  | Methyl 12-hydroxy-9-octadecenoate          |
| 10    | 26.656 | 10916390  | 9.67   | 11-Eicosenoic acid, methyl ester           |
| 11    | 26.866 | 3575666   | 3.17   | Eicosanoic acid, methyl ester              |
| 12    | 27.279 | 951745    | 0.84   | 9-Octadecenamide, (Z)-                     |
| 13    | 27.871 | 921182    | 0.82   | Phenol, 2,2'-methylenebis[6-(1,1-dimethyl  |
| 14    | 28.422 | 5871103   | 5.20   | 13-Docosenoic acid, methyl ester           |
| 15    | 28.604 | 678176    | 0.60   | Docosanoic acid, methyl ester              |
| 16    | 29.358 | 13497165  | 11.95  | .beta.-Sitosterol                          |
|       |        | 112921068 | 100.00 |  |

Major components are discussed below:

#### Methyl 12-hydroxy-9-octadecenoate (17.37%)

In Fig. 2, the peak at m/z294 (R.T.26.602) in total ion chromatogram corresponds M+[C21H42O2] +. The peak at m/z263 accounts for loss of a methoxyl.

#### 9,12,15-Octadecatrienoic acid (14.98%)

The mass spectrum of 9,12,15-octadecatrienoic acid is displayed in Fig. 3. The peak at m/z 292, which appeared at R.T.24.726 in total ion chromatogram, corresponds M+ [C19H32O2] +. The loss of a methyl function is testified by a peak at m/z277.

#### 9-Octadecenoic acid methyl ester (12.84%)

The peak at m/z 296(Fig.4), which appeared at R.T.24.702, corresponds M+ [C19H36O2] +, while the signal at m/z 264 is attributed to loss of a methoxyl function [6].

#### β-Sitosterol (11.95%)

The EI mass spectrum of β-sitosterol. The peak at m/z414, which appeared at R.T. 29.358 in the total ion chromatogram, corresponds M+ [C29H50O] +. The loss of a methyl group is demonstrated by a peak at m/z 299.

#### 9,12-Octadecadienoic acid methyl ester (10.15%)

The EI mass spectrum of 9,12-octadecadienoic showed m/z294 for M+ [C19H34O2] +. The signal at m/z 263 is due to loss of a methoxyl.

#### 11-Eicosenoic acid (9.67%)

The peak at m/z324, which appeared (Fig.7) at R.T. 26.656, corresponds M+[C21H40O2] +, while the signal at m/z292 accounts for loss of a methoxyl group.

#### Hexadecanoic acid (8.63%)

The EI mass spectrum of hexadecanoic acid. The peak at m/z 270 (R.T. 22.720) in the total ion chromatogram corresponds M+ [C17H34O2] +. The signal at m/z 239 is attributed to loss of a methoxyl.

#### 13-Docosenoic acid (5.20%)

The peak which appeared at m/z352 (R.T.28.422)

corresponds to M+[C23H44O2] +, while the signal at m/z320 corresponds to loss of a methoxyl.

#### Antimicrobial assay

*Nigella sativa* seed oil was extracted by two different methods (A: Soxhlet and B: maceration) to evaluate the effect of heating (Soxhlet) on the antimicrobial potency of the extracted oil. The oil extracted by Soxhlet was partially active against *Escherichia coli* at 5µg/ml, active at concentrations of 10 and very active at 50 µg/ml while the macerated sample was merely partially active at 50µg/ml. A partial activity was observed for *Klebsiella pneumonia* when using the Soxhlet sample at 5, 10 and 50 µg/ml, while the macerated sample was inactive at 5 µg/ml. For *Acinetobacter baumannii*, the macerated sample showed partial activity at 10 and 50 µg/ml while the Soxhlet sample was devoid of activity at all test concentrations. Noteworthy that the macerated sample gave excellent activity against the fungus *Candida albicans* at 50µg/ml, while the Soxhlet sample was devoid of activity at all tested concentrations. It seems that the potency of such extracts depends largely on the type of test organism and the concentration of the sample. Similar trends were observed for the minimum inhibition concentration [8, 9].

#### REFERENCES

1. Ncube NS, Afolayan AJ, Okoh AI, 2008. Assessment techniques of antimicrobial properties of natural compounds of plant origin: current methods and future trends. *Afr J Biotechnol.* 7, Pages 1797-1806.
2. Ramadan MF, Morsel JT, 2003. Analysis of glycolipids from black cumin (*Nigella sativa*), coriander (*Coriandrum sativum* L.) and niger (*Guizotia abyssinica* Cass.) oil seeds. *Food Chemistry.* 80, Pages 197-204.
3. Ali BH, Blunden G, 2003. Pharmacological and toxicological properties of *Nigella sativa*. *Phytother. Res.* 17, Pages 299-305.
4. El-Din K, El-Tahir H, Bakeet DM, 2006. The Black Seed *Nigella sativa* Linnaeus - A mine for multi cures: A plea for urgent clinical evaluation of its volatile oil. *J Taibah Univ. Med. Sci.* 1, Pages 1-19.
5. Randhawa MA, Alghamdi MS, 2011. Anticancer Activity of *Nigella sativa* (Black Seed). *Am J Chin Med.* 39, Pages 1075-1091.
6. Ramadan MF, 2007. Nutritional value, functional properties and nutraceuticals applications of black cumin (*Nigella sativa* L.): an overview. *Int J Food Sci Tech.* 42, Pages 1208-1218.
7. Ullah H, Rauf A, Ullah Z, et al, 2014. Density functional theory and phytochemical study of Pistagremic acid. *Acta A: Mol. Biomol. Spectrosc.* 118, Pages 210-214.